## P2656

# A Novel Chromogenic Culture Media (CHROMagar COL-APSE) for the Isolation and Differentiation of Colistin Resistant Gram-negative Pathogens 🤷 Queen Mary Muhd Haziq F Abdul Momin<sup>1</sup>, David C Bean<sup>2</sup>, Rene S Hendriksen<sup>3</sup>, Marisa Haenni<sup>4</sup>, Lynette M Phee<sup>1,5</sup>, David W Wareham<sup>1,5</sup>

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## INTRODUCTION

- Polymyxin E (colistin) and B are increasingly used as antimicrobials in the treatment of multi-drug resistant bacterial infections.
- Polymyxin resistance (PR), although intrinsic in Gram-positive and some Gram-negative species (Proteus, Morganella, Serattia spp), is now a problem in a number of other pathogens (Acinetobacter baumannii, Pseudomonas aeruginosa, Escherichia coli, Salmonella enterica, Klebsiella pneumoniae).
- Resistance arises due to mutations / insertions in genes involved in LPS biosynthesis (Ipx, pmrA/B, mgrB, phoP/Q) and / or the acquisition of phosphoethanolomine transferases (PEtN). • Of great concern is the recently described plasmid-encoded PEtN, MCR-1, now found worldwide in a
- range of animal, human and environmental bacterial isolates

## AIM

To develop a new chromogenic culture media and evaluate its sensitivity and specificity in the detection of polymyxin-resistant pathogens.

## MATERIALS AND METHODS

#### **Media Preparation**

- CHROMagar COL-APSE plates were prepared in-house using dehydrated CHROMagar base media (CHROMagar<sup>™</sup>, Paris, France) supplemented with CHROMagar COL-*APSE* supplement (Figure 1).
- This contains antimicrobials specifically selected to avoid potentiation (colistin sulfate oxazolidonone) at concentrations designed to enhance the growth of PR Gram-negative species, and suppress the growth of Gram-positive bacteria.
- CHROMagar COL-APSE media for use with samples containing Proteus spp were prepared with the addition of 50 mg/L p-nitrophenyl glycerol (PNPG).

### Bacterial Isolates and Determination of Polymyxin Minimum Inhibitory Concentration (MICs)

- Eighty-four isolates were used in the evaluation of CHROMagar COL-APSE media. • 8 isolates with intrinsic COL resistance (1 clinical human and 7 type strains), 13 COL susceptible isolates (6 clinical human, 3 clinical veterinary and 4 type strains) and 63 isolates with acquired COL
- resistance (5 clinical human and 58 clinical veterinary) contained within our collections. • The MICs of colistin (COL) and polymyxin B (POL) (0.006 – 256 µg/ml) were first determined by agar
- dilution on Mueller-Hinton II agar using a multi-point inoculator. • Susceptibility ( $\leq 2-4 \ \mu g/m$ ) and resistance (> 2-8  $\mu g/m$ ) to either COL or POL was interpreted
- according to current CLSI and / or EUCAST species specific breakpoints.

#### MCR1/2 Detection

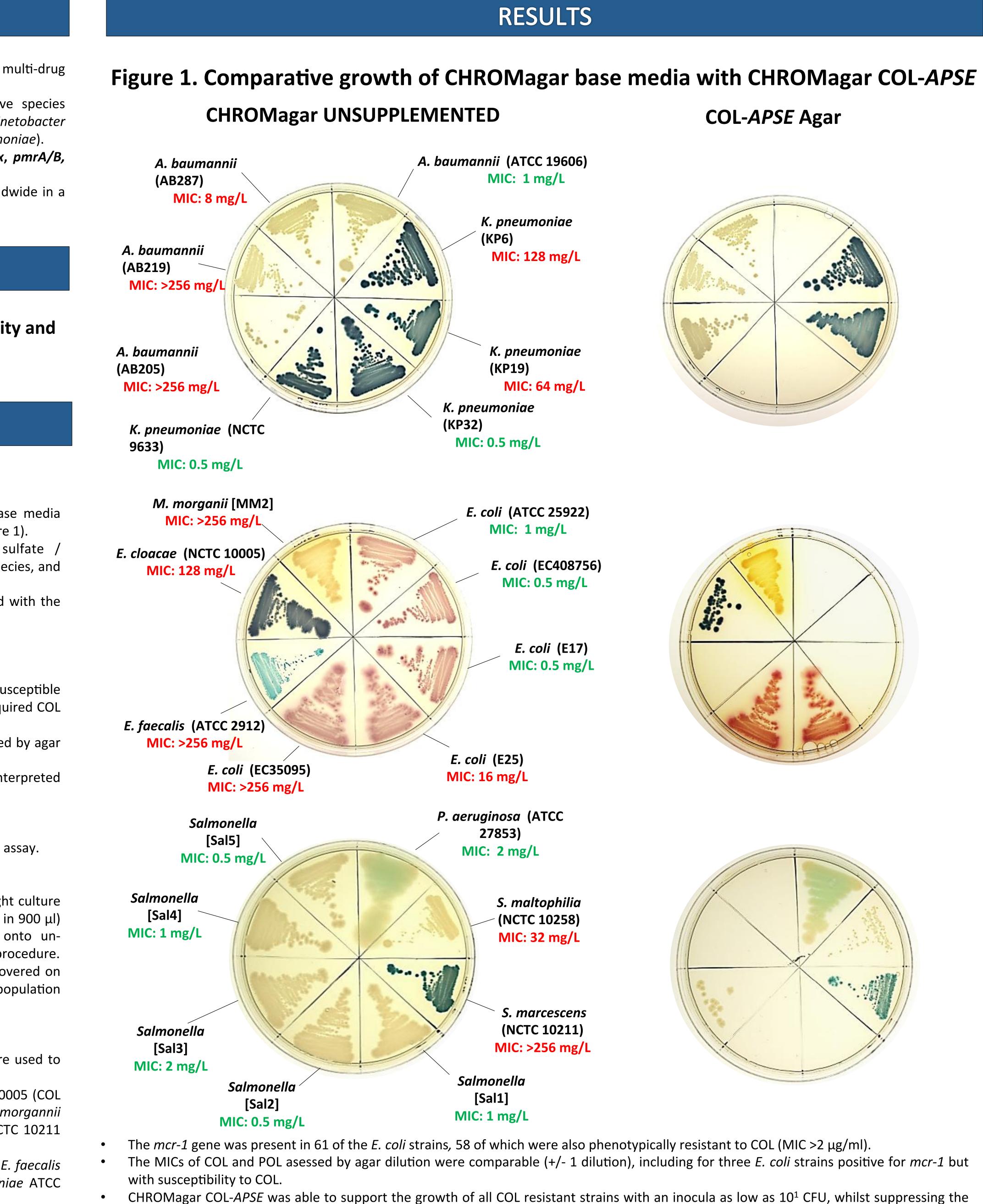
• All clinical isolates were screened for the presence of the *mcr-1/2* genes by a multiplex PCR assay.

#### Lower Limit of Detection (LLD)

- The lowest limit of detection was assessed using serial dilutions  $(10^{-1} 10^{-9})$  of an overnight culture grown at 37 °C for 24 h in 3 ml of Luria-Bertani (LB) broth. Ten-fold serial dilutions (100 μl in 900 μl) were made in phosphate-buffered saline (PBS) and 20 µl of each dilution plated onto unsupplemented MH 2 (control) and CHROMagar COL-APSE plates using the Miles and Misra procedure.
- Colony counts obtained on CHROMagar COL-APSE were subtracted from the number recovered on MH 2 agar to quantify the total number of COL resistant organisms (CFU) within the total population plated, required for viable growth on each selective media (Table 1).

#### Isolation and Differentiation of Organisms in Mixed Culture

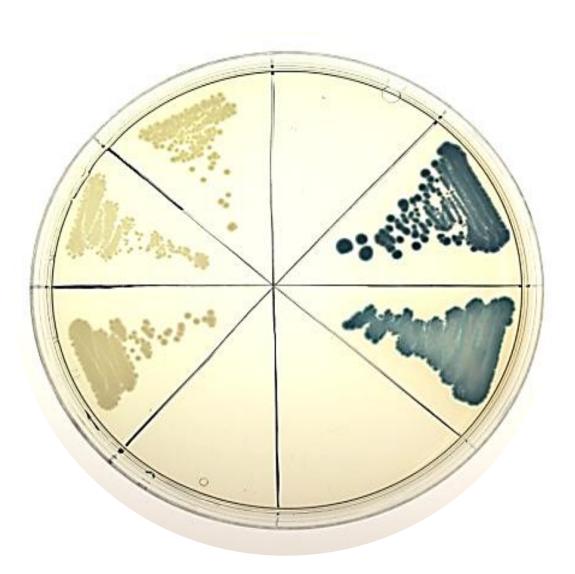
- Two pools containing mixtures of COL resistant (R) and COL susceptible (S) organisms were used to assess the performance of the media with complex polymicrobial samples (Figure 2).
- Pool 1 A. baumannii AB205 (COL resistant), E. coli E7 (COL resistant), E. cloacae NCTC 10005 (COL resistant), E. faecalis ATCC 2912 (COL resistant), K. pneumoniae KP19 (COL resistant), M. morgannii MM2 (COL resistant), P. aeruginosa ATCC 27853 (COL heteroresistant), S. marcescens NCTC 10211 (COL resistant), *P. mirabilis* NCTC 13376 (COL resistant)
- Pool 2 A. baumannii ATCC 19606 (COL susceptible), E. coli ATCC 25922 (COL susceptible), E. faecalis ATCC 2912 (COL resistant), P. aeruginosa ATCC 27853 (COL heteroresistant), K. pneumoniae ATCC 9633 (COL susceptible)
- Internal and external quality control of the stability of the media was assessed using *P. mirabilis* NCTC 13376, E. coli ATCC 25922, K. pneumoniae ATCC 9633 and E. faecalis ATCC 2912. Media for use with pools containing *P. mirabilis* NCTC 13376 was supplemented with PNPG.

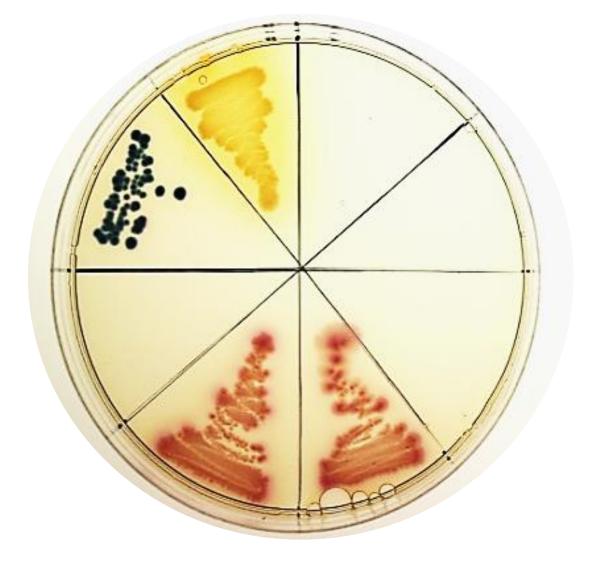


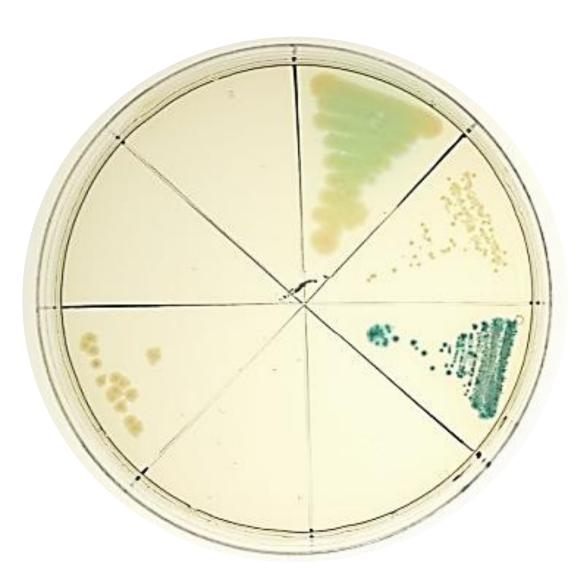
suppressing swarming of colistin resistant *Proteus spp* without affecting the performance of the media. Proteus swarming (Proteus mirabilis NCTC 13376) was reduced significantly with the addition of PNPG (Figure 2B). No growth was seen in sensitive bacterial pools +/- *Proteus mirabilis* NCTC 13376 and +/- PNPG (Figure 2C).

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# COL-APSE Agar





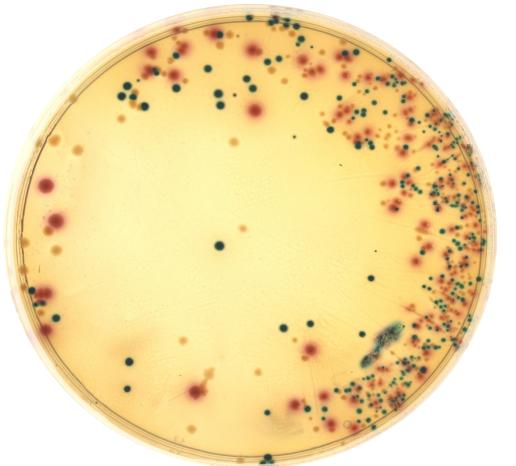


growth of all COL susceptible Gram-negative and COL resistant Gram-positive species. Notably, addition of PNPG was beneficial in

## Table 1. Lower Limit of Detection Isolates were provided by \*Denmark and #France

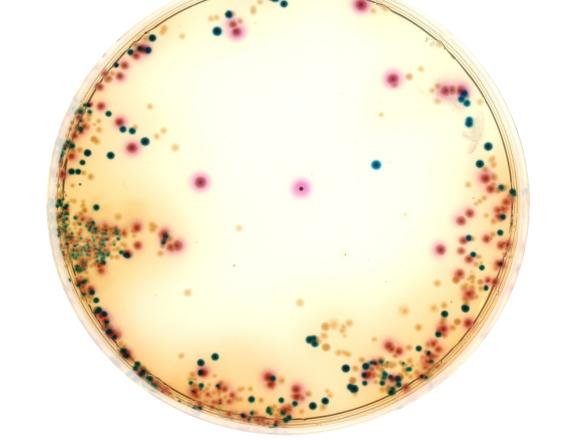
ISOLATE			Polymyxin Resistance	Lowest Limit of Detection (CFU)
	MIC (mg/L)/ A		Mechanism	
	Colistin	Polymxyin B		CHROMAgar COL-APSE
Intrinsic Resistance to Polymyxins				
<i>P. mirabilis</i> NCTC 13376	>256	>256	Intrinsic	10 1
S. marcescens NCTC 10211	>256	>256	Intrinsic	10 1
M. morgannii MM2	>256	>256	Intrinsic	10 1
<i>E. faecalis</i> ATCC 2912	>256	256	Intrinsic	>10 <sup>9</sup>
E. gallinarum ATCC 49573	>256	>256	Intrinsic	>10 8
C. albicans ATCC 10231	256	256	Intrinsic	>10 <sup>7</sup>
<i>E. cloacae</i> NCTC 10005	128	256	Intrinsic	10 <sup>2</sup>
<i>S. maltophilia</i> NCTC 10258	8	64	Intrinsic	10 1*
Susceptible to Polymyxins				
P. aeruginosa ATCC 27853	2	2	NA	10 <sup>2</sup>
Salmonella Group D (non-Typhi) Sal3	2	2	N/A	10 4
<i>E. coli</i> E17		2	MCR-1	10 <sup>7</sup>
Salmonella enterica subsp. diarizonae Sal1	1	2	N/A	10 5
A. baumannii ATCC 19606	1	2	NA	10 <sup>6</sup>
E. coli E44	1	1	MCR-1	>10 9
Salmonella enterica subsp. diarizonae Sal4	1	1	N/A	10 <sup>5</sup>
<i>K. pneumoniae</i> KP32	0.5	1	NA	10 <sup>6</sup>
Salmonella enterica subsp. diarizonae Sal2	0.5	1	N/A	10 <sup>5</sup>
Salmonella Group D (non-Typhi) Sal5	0.5	1	N/A	10 <sup>3</sup>
<i>E. coli</i> ATCC 25922	0.5	0.5	NA NA	10 <sup>6</sup>
<i>E. coli</i> 40875	0.5	0.5	MCR-1	10 <sup>6</sup>
<i>K. pneumoniae</i> ATCC 9633				>10 9
-	0.5	0.5	NA	~10
Acquired Resistance to Polymxins	0.5.0	050		101
A. baumannii AB219	>256	>256	Unknown	10 <sup>1</sup>
A. baumannii AB205	>256	32	Unknown	10 <sup>1</sup>
A. baumannii AB287	8	4	Unknown	10 1
K. pneumoniae KP6	128	256	Unknown	10 <sup>1</sup>
K. pneumoniae KP19	64	64	Unknown	10 1
<sup>#</sup> E. coli 35095	>256	>256	MCR-1	10 1
<sup>#</sup> E. coli 35175	64	32	MCR-1	10 1
<i><sup>#</sup>E. coli</i> (n=4)	32	32	MCR-1	10 <sup>1</sup>
* <i>E. coli</i> (n=12)	32	16	MCR-1	10 1*
* <i>E. coli</i> E27	16	32	MCR-1	10 <sup>1</sup>
* <sup>#</sup> <i>E. coli</i> (n=34)	16	16	MCR-1	10 <sup>1</sup>
* <sup>#</sup> <i>E. coli</i> (n=3)	16	8	MCR-1	10 <sup>1</sup>
* <i>E. coli</i> E45	8	16	MCR-1	10 <sup>1</sup>
<sup>#</sup> E. coli 32218	8	8	MCR-1	10 <sup>1</sup>

## Figure 2. Simulated bacterial pools inoculated using L-spreader method



A) CHROMagar COL-APSE

- Chemother 59:2780-2784.



B) CHROMagar COL-APSE + 50 mg/L pnitrophenyl glycerol (PNPG)



C) CHROMagar COL-APSE +/- 50 mg/L p-nitrophenyl glycerol (PNPG)

## CONCLUSION

• CHROMagar COL-APSE is a sensitive and specific media for the growth of COL resistant bacterial pathogens with a lower limit of detection of 10<sup>1</sup> CFU

• CHROMagar COL-APSE may be useful as a primary isolation media in the surveillance and recovery of COL resistant bacteria from complex human, veterinary and environmental samples especially those with plasmid mediated MCR-1 or novel mechanisms of polymyxin resistance.

## REFERENCES

1. Liu Y-Y, Wang Y, Walsh TR, Yi L-X, Zhang R, Spencer J, Doi Y, Tian G, Dong B, Huang X, Yu L-F, Gu D, Ren H, Chen X, Lv L, He D, Zhou H, Liang Z, Liu J-H, Shen J. 2016. Emergence of plasmidmediated colistin resistance mechanism MCR-1 in animals and human beings in China: a microbiological and molecular biological study. Lancet Infect Dis 16:161–168. 2. Jayol A, Nordmann P, Brink A, Poirel L. 2015. Heteroresistance to colistin in Klebsiella pneumoniae associated with alterations in the PhoPQ regulatory system. Antimicrob Agents

3. Agersø Y, Torpdahl M, Zachariasen C, Seyfarth A, Hammerum AM, Nielsen EM. 2012. Tentative colistin epidemiological cut-off value for Salmonella spp. Foodborne Pathog Dis 9:367-369.