

## Replacement of *Candida albicans* with *C. dubliniensis* in Human Immunodeficiency Virus-Infected Patients with Oropharyngeal Candidiasis Treated with Fluconazole

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*Candida dubliniensis* is an opportunistic yeast that has been increasingly implicated in oropharyngeal candidiasis (OPC) in human immunodeficiency virus (HIV)-infected patients but may be underreported due to its similarity with *Candida albicans*. Although most *C. dubliniensis* isolates are susceptible to fluconazole, the inducibility of azole resistance in vitro has been reported. Thus, the use of fluconazole prophylaxis in the treatment of these patients may have contributed to the increasing rates of isolation of *C. dubliniensis*. In this study, yeast strains were collected from the oral cavities of HIV-infected patients enrolled in a longitudinal study of OPC. Patients received fluconazole for the suppression or treatment of OPC, and isolates collected at both study entry and end of study were chosen for analysis. Samples were plated on CHROMagar *Candida* medium for initial isolation and further identified by Southern blot analysis with the species-specific probes Ca3 (for *C. albicans*) and Cd25 (for *C. dubliniensis*). Fluconazole MICs were determined by using NCCLS methods. At study entry, susceptible *C. albicans* isolates were recovered from oral samples in 42 patients who were followed longitudinally (1 to 36 months). *C. albicans* strains from 12 of these patients developed fluconazole resistance (fluconazole MIC,  $\geq 64$   $\mu\text{g/ml}$ ). *C. dubliniensis* was not detected at end of study in any of these patients. Of the remaining 30 patients, eight (27%) demonstrated a replacement of *C. albicans* by *C. dubliniensis* when a comparison of isolates obtained at baseline and those from the last culture was done. For the 22 of these 30 patients in whom no switch in species was detected, the fluconazole MICs for initial and end-of-study *C. albicans* isolates ranged from 0.125 to 2.0  $\mu\text{g/ml}$ . For the eight patients in whom a switch to *C. dubliniensis* was detected, the fluconazole MICs for *C. dubliniensis* isolates at end of study ranged from 0.25 to 64  $\mu\text{g/ml}$ ; the fluconazole MICs for isolates from six patients were 0.25 to 2.0  $\mu\text{g/ml}$  and those for the other two were 32 and 64  $\mu\text{g/ml}$ , respectively. In conclusion, a considerable number of patients initially infected with *C. albicans* strains that failed to develop fluconazole resistance demonstrated a switch to *C. dubliniensis*. *C. dubliniensis* in this setting may be underestimated due to lack of identification and may occur due to the impact of fluconazole on the ecology of oral yeast species.

*Candida dubliniensis* is a causative agent of oropharyngeal candidiasis (OPC) in human immunodeficiency virus (HIV)-infected and AIDS patients (10, 34; D. C. Coleman, D. J. Sullivan, and J. M. Mossman, Letter, J. Clin. Microbiol. **35**: 3011–3012). It is a newly identified species of *Candida*, phenotypically similar to (chlamydospore and germ tube positive), but genetically distinct from, *Candida albicans* (1, 35, 36). Indeed, *C. dubliniensis* was initially difficult to distinguish from *C. albicans* and was often misidentified as such in standard clinical laboratory tests, because it shares phenotypic characteristics with *C. albicans* (10, 31, 34, 35). Thus, due to the phenotypic similarity of the two organisms, there is a growing body of evidence suggesting that many previous cases of OPC thought to have been caused by *C. albicans* were in reality caused by *C. dubliniensis* (6, 10, 31). Since its identification as an etiologic agent of OPC, *C. dubliniensis* has received increasing attention.

Although it was initially implicated in OPC in HIV-infected patients, particularly those with recurrent infections, *C. dubliniensis* has since been found to be both an oral carriage organism and a causative agent of oral candidiasis in HIV-negative individuals (1–2, 5, 10, 12–14, 21, 24, 25, 29, 32–35; Coleman et al., letter). The interaction and ecological relationships of different *Candida* species in the oropharynx are not completely understood and appropriately constitute a topic of active research. In a recent study that examined the growth competition between *C. albicans* and *C. dubliniensis* under broth and biofilm growth conditions, it was shown that *C. albicans* had a competitive advantage over *C. dubliniensis* (9). The clinical significance of this organism is yet to be determined, but it has been demonstrated that, contrary to fluconazole (FLU) resistance in *C. albicans*, FLU resistance in *C. dubliniensis* is easily inducible in vitro (15). Thus, prolonged use of FLU in HIV-infected patients may impact the epidemiology of OPC.

The present study was undertaken to further define the complex microbial ecology in the oropharynxes of HIV-infected patients exposed to FLU for the treatment of OPC and to increase our understanding of the role of *C. dubliniensis* in OPC.

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## MATERIALS AND METHODS

**Clinical samples.** Yeast isolates were recovered from swabs and/or oral rinses taken from the oropharynxes of HIV-infected patients enrolled in a longitudinal study to assess the development of FLU resistance. Sixty-four patients were enrolled and monitored clinically for the presence of OPC. Cultures of the oropharynx were obtained at the time of enrollment and either during times of clinical OPC or at quarterly intervals if OPC was not present. All patients were treated initially with FLU at a dose of 100 mg/day. Doses of FLU were increased up to 800 mg/day in those individuals who developed clinical resistance, which is defined as the clinical requirement for increasing FLU doses for response (10, 26–28). All oral samples were screened with the chromogenic medium CHROMagar *Candida* (CHROMagar Company, Paris, France) (17, 18) and initially identified as *C. albicans* by colony color and a positive germ tube test in serum. Selected yeast isolates included those obtained on both initial and final cultures from these patients.

**Susceptibility testing.** *Candida* isolates were tested for susceptibility to FLU in accordance with NCCLS methods (16). *Candida* isolates were considered susceptible (MIC, 8 µg/ml), susceptible dose dependent (MIC, 16 to 32 µg/ml), or resistant (MIC, ≥64 µg/ml). All antifungal susceptibility testing was performed at the Fungal Testing Laboratory, University of Texas Health Science Center San Antonio, San Antonio.

**DNA typing techniques.** Electrophoretic karyotype and fingerprinting analyses of *EcoRI*-digested chromosomal DNA with the repetitive probes Ca3, specific for *C. albicans* (30), and Cd25, specific for *C. dubliniensis* (8), were performed as previously described (10, 11, 19). Hybridization with probes Ca3 and Cd25 was also used as a genotypic tool to confirm the identity of the isolates as *C. albicans* or *C. dubliniensis*, respectively. In brief, chromosomal DNA from each isolate was prepared in agarose plugs and separated by pulsed-field gel electrophoresis (Bio-Rad, Hercules, Calif.). Restriction fragment length polymorphism patterns were obtained by digestion of chromosomal DNA by using *SfiI* and *EcoRI* (Boehringer-Mannheim, Indianapolis, Ind.). Digested DNA present in the restriction fragment length polymorphism gels was transferred to nylon membranes (Nytran; Schleicher and Schuell, Keene, N.H.) and hybridized with probe Ca3 and subsequently with probe Cd25 (after removal of the bound probe Ca3), radioactively labeled by random priming (Random Primers DNA labeling system; GibcoBRL, Gaithersburg, Md.), and exposed to autoradiography film (Du Pont, Wilmington, Del.) overnight at –70°C.

## RESULTS

Of the 64 patients enrolled in the study, initial cultures of the oropharynx specimens from 42 demonstrated FLU-susceptible *C. albicans* isolates. Of these 42 patients, 12 developed FLU-resistant (FLU MIC, ≥64 µg/ml) *C. albicans* on their final cultures (species identity for all resistant isolates was confirmed at the DNA level) while 30 patients did not. Upon further evaluation of all final isolates from these 30 patients, eight patients whose initial cultures had demonstrated a FLU-susceptible *C. albicans* isolate showed a change to a *C. dubliniensis* isolate at end of study, as shown in Table 1. *C. dubliniensis* isolates were susceptible to FLU at MICs ranging from 0.25 to 64 µg/ml. The FLU MICs for six isolates were 0.25 to 2.0 µg/ml, while those for two other isolates were 32 and 64 µg/ml, respectively.

Strain delineation of isolates was performed by electrophoretic karyotype analysis, as shown in Fig. 1. All initial isolates demonstrated chromosomal banding patterns consistent with that of *C. albicans*. These patterns, however, were distinctly different from those obtained for the end-of-study, or final, isolates. Final isolates (Fig. 1, lanes B) all demonstrated a distinct low-molecular-weight band that was not found for any of the initial isolates, which is characteristic of *C. dubliniensis*. Fingerprinting analysis with the probes specific for *C. albicans* (Ca3) and *C. dubliniensis* (Cd25) are shown in Fig. 2 and 3, respectively. As shown in Fig. 2, all initial isolates hybridized with Ca3, as demonstrated by autoradiography, while

TABLE 1. Initial and end-of-study culture isolates from 30 patients and their FLU susceptibilities

Patient no.	Culture results			
	Initial		End of study	
	Isolate (no.)	FLU MIC (µg/ml)	Isolate (no.)	FLU MIC (µg/ml)
2	<i>C. albicans</i> (321)	0.25	<i>C. dubliniensis</i> (1884)	32
4	<i>C. albicans</i> (346)	0.25	<i>C. albicans</i> (1421)	0.25
6	<i>C. albicans</i> (389)	2.0	<i>C. albicans</i> (6261)	0.5
8	<i>C. albicans</i> (434)	2.0	<i>C. albicans</i> (944)	0.5
10	<i>C. albicans</i> (442)	0.5	<i>C. albicans</i> (476)	0.5
12	<i>C. albicans</i> (508)	0.25	<i>C. albicans</i> (1384)	0.25
13	<i>C. albicans</i> (532)	1.0	<i>C. albicans</i> (680)	0.5
19	<i>C. albicans</i> (730)	0.5	<i>C. albicans</i> (5754)	0.25
20	<i>C. albicans</i> (745)	0.5	<i>C. albicans</i> (796)	2.0
21	<i>C. albicans</i> (801)	2.0	<i>C. dubliniensis</i> (1955)	0.25
23	<i>C. albicans</i> (958)	2.0	<i>C. albicans</i> (1932)	4.0
24	<i>C. albicans</i> (1017)	0.5	<i>C. albicans</i> (2927)	2.0
25	<i>C. albicans</i> (1032)	0.25	<i>C. dubliniensis</i> (3890)	0.5
26	<i>C. albicans</i> (1148)	0.5	<i>C. dubliniensis</i> (3651)	2.0
29	<i>C. albicans</i> (1224)	1.0	<i>C. albicans</i> (1576)	1.0
31	<i>C. albicans</i> (1297)	0.25	<i>C. albicans</i> (1355)	1.0
32	<i>C. albicans</i> (1401)	0.5	<i>C. dubliniensis</i> (2162)	0.5
33	<i>C. albicans</i> (1454)	0.125	<i>C. dubliniensis</i> (5579)	0.5
36	<i>C. albicans</i> (1538)	0.125	<i>C. albicans</i> (3721)	0.5
38	<i>C. albicans</i> (1483)	0.25	<i>C. albicans</i> (1609)	0.25
39	<i>C. albicans</i> (1486)	0.5	<i>C. albicans</i> (6074)	1.0
40	<i>C. albicans</i> (1490)	0.5	<i>C. albicans</i> (2511)	4.0
47	<i>C. albicans</i> (1922)	1.0	<i>C. albicans</i> (3078)	16
52	<i>C. albicans</i> (2325)	1.0	<i>C. albicans</i> (4424)	0.5
53	<i>C. albicans</i> (2352)	0.5	<i>C. albicans</i> (5842)	2.0
55	<i>C. albicans</i> (2531)	0.5	<i>C. albicans</i> (6169)	0.5
56	<i>C. albicans</i> (3840)	4.0	<i>C. albicans</i> (4732)	4.0
62	<i>C. albicans</i> (2925)	0.5	<i>C. albicans</i> (3315)	0.5
63	<i>C. albicans</i> (3070)	1.0	<i>C. dubliniensis</i> (3152)	0.25
64	<i>C. albicans</i> (4018)	1.0	<i>C. dubliniensis</i> (4380)	64

none of the final isolates demonstrated hybridization activity. In Fig. 3, however, the converse is true. None of the initial isolates hybridized with Cd25, while each of the final isolates demonstrated reactivity. These hybridization patterns showed that all initial isolates were *C. albicans* and all final isolates were *C. dubliniensis*.

## DISCUSSION

The increased use of antifungal agents (mainly FLU) may be responsible for the development of more highly resistant microorganisms and may impact the ecology of oral yeast species by effecting a shift toward species with innate resistance or those that are more prone to develop resistance to drugs such as FLU. Although most *C. dubliniensis* clinical isolates are susceptible to azole derivatives (2, 10, 15, 20, 22), our group and others have reported the isolation of FLU-resistant *C. dubliniensis* clinical isolates from the oral cavities of HIV-infected patients with OPC and prior exposure to FLU (10, 15). In addition, derivatives exhibiting a stable FLU-resistant phenotype can be readily generated in vitro from FLU-susceptible isolates (15), a phenomenon that is not easily observed with *C. albicans*, thus indicating that *C. dubliniensis* may display a higher propensity to develop azole resistance than *C. albicans*.

Our results indicate that replacement of *C. albicans* by *C. dubliniensis* occurred in a considerable number (27%) of pa-

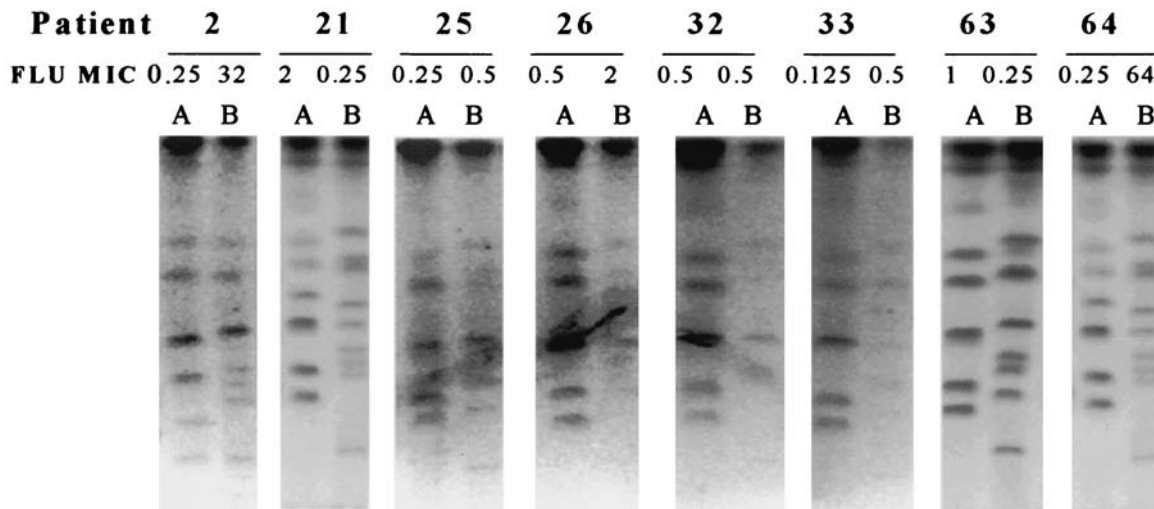


FIG. 1. Electrophoretic karyotype analysis results for *Candida* isolates from eight patients in whom the culture isolate changed from *C. albicans* to *C. dubliniensis*. Patient numbers are given at the top of the figure. Lanes: A, initial isolates; B, final isolates. FLU MICs are in micrograms per milliliter.

tients treated with FLU who failed to develop FLU-resistant *C. albicans*. Of note, virtually all of these isolates were initially identified incorrectly as *C. albicans* by the phenotypic methods routinely used in our laboratory, and only subsequent use of genotypic methods (karyotyping and Cd25 fingerprinting) allowed correct identification of the isolates as *C. dubliniensis*. For the patients in the present study, the FLU MICs for initial and end-of-study *C. albicans* isolates from those who did not exhibit a change in species ( $n = 22$ ) ranged from 0.125 to 2.0  $\mu\text{g/ml}$  whereas the FLU MICs for *C. dubliniensis* isolates from end-of-study cultures for eight patients ranged from 0.25 to 64  $\mu\text{g/ml}$ . Interestingly, replacement of *C. albicans* by *C. dubliniensis* was not observed in patients whose *C. albicans* strains were able to develop FLU resistance. These results suggest that the antifungal pressure exerted by prolonged FLU treatment influences the oral microbial ecology in these patients.

Species that are better able to adapt to the antifungal pressure may persist over those that are effectively suppressed by the treatment. However, persistence of oral microorganisms as commensal or infectious organisms may also depend on other factors not related to resistance. Indeed, the majority of *C. dubliniensis* isolates recovered at the end of study did not demonstrate decreased susceptibility to FLU (Table 1). We have previously demonstrated that, in the absence of antifungal pressure, *C. albicans* predominates over *C. dubliniensis* in normal growth competition assays. However, adherent populations of *C. dubliniensis* were able to withstand the competitive pressure of *C. albicans* (9). Some factors that may also favor the survival of *C. dubliniensis* in the oral cavity are its intrinsic cell surface hydrophobicity, its ability to bind to oral bacteria, and its ability to form biofilms (3, 4, 7, 23). The presence of subinhibitory concentrations of FLU may alter

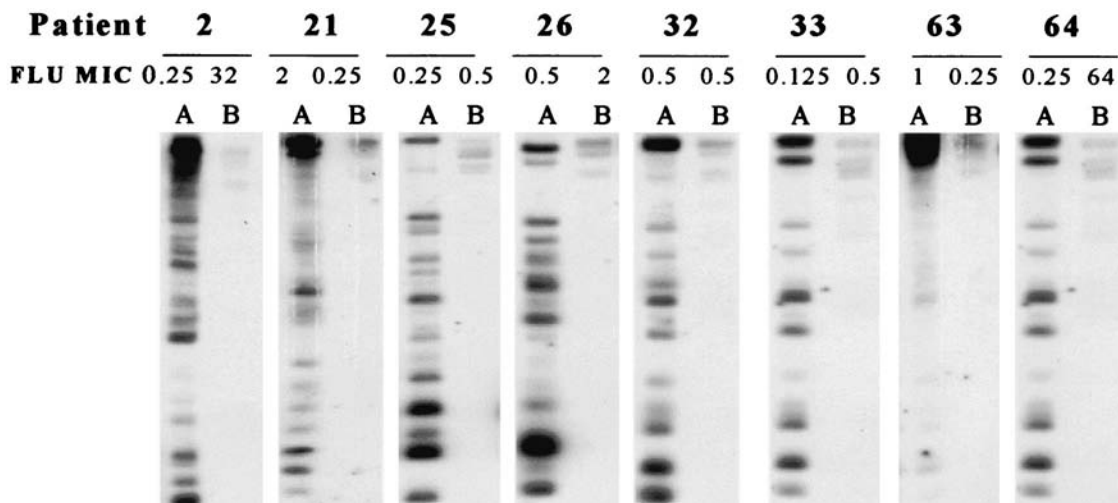


FIG. 2. DNA fingerprinting analysis of *EcoRI*-digested chromosomal DNA with the radiolabeled Ca3 probe, which is specific for *C. albicans*. Patient numbers are given at the top of the figure. Lanes: A, initial isolates; B, final isolates. FLU MICs are in micrograms per milliliter.



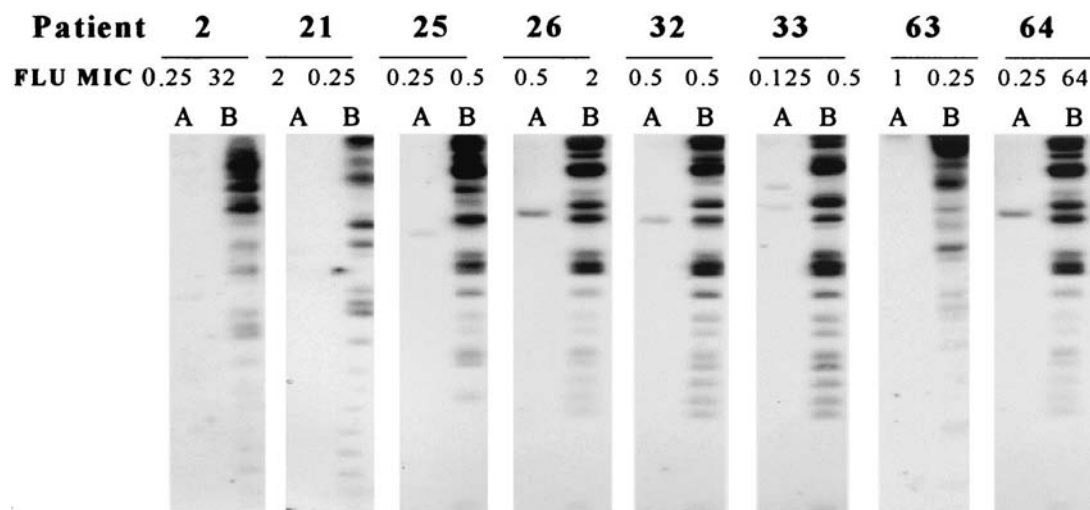


FIG. 3. DNA fingerprinting analysis of *Eco*RI-digested chromosomal DNA with the radiolabeled Cd25 probe, which is specific for *C. dubliniensis*. Patient numbers are given at the top of the figure. Lanes: A, initial isolates; B, final isolates. FLU MICs are in micrograms per milliliter.

these properties. These factors, together with better identification techniques, may be responsible for the increasing rates of isolation of this newly described species.

In summary, in our longitudinal study of FLU resistance in HIV-infected patients, for a considerable number of patients initially infected with *C. albicans* strains that failed to develop FLU resistance (more than one-fourth), later cultures showed a change in species isolated to *C. dubliniensis*. Identification of *C. dubliniensis* required the use of genotypic methods. Thus, *C. dubliniensis* in this setting may be underestimated due to lack of proper identification and may occur, in part, because of the impact of FLU therapy on the ecology of oral yeast species.

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#### REFERENCES

- Coleman, D., D. Sullivan, B. Harrington, K. Haynes, M. Henman, D. Shanley, D. Bennett, G. Moran, C. McCreary, and L. O'Neill. 1997. Molecular and phenotypic analysis of *Candida dubliniensis*: a recently identified species linked with oral candidosis in HIV-infected and AIDS patients. *Oral Dis.* 3(Suppl. 1):S96-S101.
- Coleman, D. C., M. G. Rinaldi, K. A. Haynes, J. H. Rex, R. C. Summerbell, E. J. Anaissie, A. Li, and D. J. Sullivan. 1998. Importance of *Candida* species other than *Candida albicans* as opportunistic pathogens. *Med. Mycol.* 36(Suppl. 1):156-165.
- Hazen, K. C., J. G. Wu, and J. Masuoka. 2001. Comparison of the hydrophobic properties of *Candida albicans* and *Candida dubliniensis*. *Infect. Immun.* 69:779-786.
- Jabra-Rizk, M. A., W. A. Falkler, W. G. Merz, J. I. Kelley, A. A. M. Baqui, and T. F. Meiller. 1999. *Candida dubliniensis* and *Candida albicans* display surface variations consistent with observed intergeneric coaggregation. *Rev. Iberoam. Micol.* 16:187-193.
- Jabra-Rizk, M. A., A. A. M. Baqui, J. I. Kelley, W. A. Falkler, Jr., W. G. Merz, and T. F. Meiller. 1999. Identification of *Candida dubliniensis* in a prospective study of patients in the United States. *J. Clin. Microbiol.* 37:321-326.
- Jabra-Rizk, M. A., W. A. Falkler, Jr., W. G. Merz, A. A. M. Baqui, J. I. Kelley, and T. F. Meiller. 2000. Retrospective identification and characterization of *Candida dubliniensis* isolates among *Candida albicans* clinical laboratory isolates from human immunodeficiency virus (HIV)-infected and non-HIV-infected individuals. *J. Clin. Microbiol.* 38:2423-2426.
- Jabra-Rizk, M. A., W. A. Falkler, Jr., W. G. Merz, J. I. Kelley, A. A. M. Baqui, and T. F. Meiller. 1999. Coaggregation of *Candida dubliniensis* with *Fusobacterium nucleatum*. *J. Clin. Microbiol.* 37:1464-1468.
- Joly, S., C. Pujol, M. Rysz, K. Vargas, and D. R. Soll. 1999. Development and characterization of complex DNA fingerprinting probes for the infectious yeast *Candida dubliniensis*. *J. Clin. Microbiol.* 37:1035-1044.
- Kirkpatrick, W. R., J. L. Lopez-Ribot, R. K. McAtee, and T. F. Patterson. 2000. Growth competition between *Candida dubliniensis* and *Candida albicans* under broth and biofilm growing conditions. *J. Clin. Microbiol.* 38:902-904.
- Kirkpatrick, W. R., S. G. Revankar, R. K. McAtee, J. L. Lopez-Ribot, A. W. Fothergill, D. I. McCarthy, S. E. Sanche, R. A. Cantu, M. G. Rinaldi, and T. F. Patterson. 1998. Detection of *Candida dubliniensis* in oropharyngeal samples from human immunodeficiency virus-infected patients in North America by primary CHROMagar *Candida* screening and susceptibility testing of isolates. *J. Clin. Microbiol.* 36:3007-3012.
- López-Ribot, J. L., R. K. McAtee, W. R. Kirkpatrick, and T. F. Patterson. 2000. Comparison of DNA-based typing methods to assess genetic diversity and relatedness among *Candida albicans* clinical isolates. *Rev. Iberoam. Micol.* 17:49-54.
- McCullough, M. J., K. V. Clemons, and D. A. Stevens. 1999. Molecular and phenotypic characterization of genotypic *Candida albicans* subgroups and comparison with *Candida dubliniensis* and *Candida stellatoidea*. *J. Clin. Microbiol.* 37:417-421.
- Meiller, T. F., M. A. Jabra-Rizk, A. Baqui, J. I. Kelley, V. I. Meeks, W. G. Merz, and W. A. Falkler. 1999. Oral *Candida dubliniensis* as a clinically important species in HIV-seropositive patients in the United States. *Oral Surg. Oral Med. Oral Pathol. Oral Radiol. Endod.* 88:573-580.
- Meis, J. F., M. Ruhnke, B. E. De Pauw, F. C. Odds, W. Siegert, and P. E. Verweij. 1999. *Candida dubliniensis* candidemia in patients with chemotherapy-induced neutropenia and bone marrow transplantation. *Emerg. Infect. Dis.* 5:150-153.
- Moran, G. P., D. J. Sullivan, M. C. Henman, C. E. McCreary, B. J. Harrington, D. B. Shanley, and D. C. Coleman. 1997. Antifungal drug susceptibilities of oral *Candida dubliniensis* isolates from human immunodeficiency virus (HIV)-infected and non-HIV-infected subjects and generation of stable fluconazole-resistant derivatives in vitro. *Antimicrob. Agents Chemother.* 41:617-623.
- National Committee for Clinical Laboratory Standards. 1997. Reference method for broth dilution antifungal susceptibility testing of yeasts. Approved standard. NCCLS document M27-A. National Committee for Clinical Laboratory Standards, Wayne, Pa.
- Patterson, T. F., W. R. Kirkpatrick, S. G. Revankar, R. K. McAtee, A. W. Fothergill, D. I. McCarthy, and M. G. Rinaldi. 1996. Comparative evaluation of macrodilution and chromogenic agar screening for determining fluconazole susceptibility of *Candida albicans*. *J. Clin. Microbiol.* 34:3237-3239.

18. Patterson, T. F., S. G. Revankar, W. R. Kirkpatrick, O. Dib, A. W. Fothergill, S. W. Redding, D. A. Sutton, and M. G. Rinaldi. 1996. Simple method for detecting fluconazole-resistant yeasts with chromogenic agar. *J. Clin. Microbiol.* **34**:1794–1797.
19. Perea, S., J. L. López-Ribot, B. L. Wickes, W. R. Kirkpatrick, O. P. Dib, S. P. Bachmann, S. M. Keller, M. Martínez, and T. F. Patterson. 2002. Molecular mechanisms of fluconazole resistance in *Candida dubliniensis* isolates from human immunodeficiency virus-infected patients with oropharyngeal candidiasis. *Antimicrob. Agents Chemother.* **46**:1695–1703.
20. Pfaller, M. A., S. A. Messer, S. Gee, S. Joly, C. Pujol, D. J. Sullivan, D. C. Coleman, and D. R. Soll. 1999. In vitro susceptibilities of *Candida dubliniensis* isolates tested against the new triazole and echinocandin antifungal agents. *J. Clin. Microbiol.* **37**:870–872.
21. Polacheck, I., J. Strahilevitz, D. Sullivan, S. Donnelly, I. F. Salkin, and D. C. Coleman. 2000. Recovery of *Candida dubliniensis* from non-human immunodeficiency virus-infected patients in Israel. *J. Clin. Microbiol.* **38**:170–174.
22. Quindos, G., A. J. Carrillo-Munoz, M. P. Arevalo, J. Salgado, R. Alonso-Vargas, J. M. Rodrigo, M. T. Ruesga, A. Valverde, J. Peman, E. Canton, E. Martín-Mazuelos, and J. Ponton. 2000. In vitro susceptibility of *Candida dubliniensis* to current and new antifungal agents. *Chemotherapy* **46**:395–401.
23. Ramage, G., K. Vande Walle, B. L. Wickes, and J. L. López-Ribot. 2001. Biofilm formation by *Candida dubliniensis*. *J. Clin. Microbiol.* **39**:3234–3240.
24. Redding, S. W., C. W. Bailey, J. L. Lopez-Ribot, W. R. Kirkpatrick, A. W. Fothergill, M. G. Rinaldi, and T. F. Patterson. 2001. *Candida dubliniensis* in radiation-induced oropharyngeal candidiasis. *Oral Surg. Oral Med. Oral Pathol. Oral Radiol. Endod.* **91**:659–662.
25. Redding, S. W., R. C. Zellars, W. R. Kirkpatrick, R. K. McAtee, M. A. Caceres, A. W. Fothergill, J. L. Lopez-Ribot, C. W. Bailey, M. G. Rinaldi, and T. F. Patterson. 1999. Epidemiology of oropharyngeal *Candida* colonization and infection in patients receiving radiation for head and neck cancer. *J. Clin. Microbiol.* **37**:3896–3900.
26. Revankar, S. G., O. P. Dib, W. R. Kirkpatrick, R. K. McAtee, A. W. Fothergill, M. G. Rinaldi, S. W. Redding, and T. F. Patterson. 1998. Clinical evaluation and microbiology of oropharyngeal infection due to fluconazole-resistant *Candida* in human immunodeficiency virus-infected patients. *Clin. Infect. Dis.* **26**:960–963.
27. Revankar, S. G., W. R. Kirkpatrick, R. K. McAtee, O. P. Dib, A. W. Fothergill, S. W. Redding, M. G. Rinaldi, S. G. Hilsenbeck, and T. F. Patterson. 1998. A randomized trial of continuous or intermittent therapy with fluconazole for oropharyngeal candidiasis in HIV-infected patients: clinical outcomes and development of fluconazole resistance. *Am. J. Med.* **105**:7–11.
28. Revankar, S. G., W. R. Kirkpatrick, R. K. McAtee, O. P. Dib, A. W. Fothergill, S. W. Redding, M. G. Rinaldi, and T. F. Patterson. 1996. Detection and significance of fluconazole resistance in oropharyngeal candidiasis in human immunodeficiency virus-infected patients. *J. Infect. Dis.* **174**:821–827.
29. Ruhnke, M., A. Schmidt-Westhausen, and J. Morschhauser. 2000. Development of simultaneous resistance to fluconazole in *Candida albicans* and *Candida dubliniensis* in a patient with AIDS. *J. Antimicrob. Chemother.* **46**:291–295.
30. Schmid, J., E. Voss, and D. R. Soll. 1990. Computer-assisted methods for assessing strain relatedness in *Candida albicans* by fingerprinting with the moderately repetitive sequence Ca3. *J. Clin. Microbiol.* **28**:1236–1243.
31. Schoofs, A., F. C. Odds, R. Colebunders, M. Ieven, and H. Goossens. 1997. Use of specialised isolation media for recognition and identification of *Candida dubliniensis* isolates from HIV-infected patients. *Eur. J. Clin. Microbiol. Infect. Dis.* **16**:296–300.
32. Schorling, S. R., H. C. Kortinga, M. Froschb, and F. A. Muhlschlegel. 2000. The role of *Candida dubliniensis* in oral candidiasis in human immunodeficiency virus-infected individuals. *Crit. Rev. Microbiol.* **26**:59–68.
33. Sullivan, D., and D. Coleman. 1997. *Candida dubliniensis*: an emerging opportunistic pathogen. *Curr. Top. Med. Mycol.* **8**:15–25.
34. Sullivan, D., and D. Coleman. 1998. *Candida dubliniensis*: characteristics and identification. *J. Clin. Microbiol.* **36**:329–334.
35. Sullivan, D. J., T. J. Westerneng, K. A. Haynes, D. E. Bennett, and D. C. Coleman. 1995. *Candida dubliniensis* sp. nov.: phenotypic and molecular characterization of a novel species associated with oral candidosis in HIV-infected individuals. *Microbiology* **141**:1507–1521.