

Study of an acid phosphatase test for the confirmation of *Clostridium perfringens* directly from CHROMagar™ C.perfringens agar medium

CONFIDENTIAL

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This report contains 15 pages, including 4 pages of appendices.
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1. Introduction

The aim of this study is to evaluate the effectiveness of an acid phosphatase test to differentiate *Clostridium perfringens* from other potentially false positive bacteria in the CHROMagar™ C.perfringens medium.

The ISO 14189:2017 standard, allowing the enumeration of *Clostridium perfringens* in water by membrane filtration, provides an acid phosphatase test for the confirmation of the characteristic colonies.

The study aims at verifying that the replacement of the FAST Blue B salt by the FAST Blue RR salt in the acid phosphatase reagent optimizes the interpretation of the test on CHROMagar™ C.perfringens agar.

2. Materials and methods

2.1. Materials

2.1.1. Devices

The devices used in this study are the ones usually encountered in a laboratory. The culture media were incubated in anaerobic jars containing anaerogenic generators (Anaerogen ThermoScientific, ref: AG0025A) and an anaerobic control (Anaerotest Merck, ref: 15112). The paper used to carry out the acid phosphatase test is white calligraph type blotting paper at a grammage of 100 g/m².

2.1.2. Culture media

The culture media used were as follows:

- Tryptone Sulfite Cycloserine agar (OXOID - ref: CM0587)
- CHROMagar™ C.perfringens agar (Base ref: PF652(B) / Supplements ref: PF652(S1) and ref: PF652(S2))
- Trypto caseine Soja Agar (BioMérieux – ref : 41466)
- Columbia 5% Sheep Blood Agar (BioMérieux – ref: 43041)

2.1.3. Reagents

The acid phosphatase reagent used was as follows:

- | | |
|--|-------|
| - 1-Naphthylphosphate monosodium salt (CAS N°81012-89-7) | 0,4 g |
| - FAST Blue RR salt (4-Benzoylamino-2,5-dimethoxybenzenediazonium chloride hemi (zinc chloride) salt) (CAS N°14726-29-5) | 0,8 g |
| - Acetate buffer (pH 4,6 ± 0,2) | 20 ml |
| - Glacial acetic acid (CAS N°64-19-7) | |
| - Sodium acetate (CAS N°127-09-3) | |
- Acetate buffer has been prepared by dissolving 0,3 ml glacial acid acetic and 0,4 g sodium acetate in deionized water qsp to 1000 ml.

The acid phosphatase reagent was obtained by dissolving the ingredients in the acetate buffer and allow to stand for 60 ± 5 min at 5 ± 3 °C to allow precipitation. The solution was passed through a fluted filter to remove the precipitate.

2.2. Methods

2.2.1. Strain culture protocol

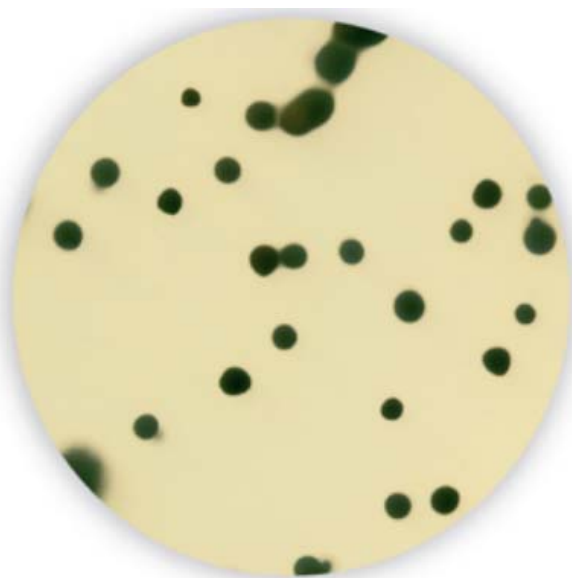
During this study, 15 strains of *Clostridium perfringens* from different origins were tested, as well as 7 *Clostridium* spp. exclusive of *Clostridium perfringens* strains. Eighteen strains which may have characteristics close to *Clostridium perfringens* on CHROMagar™ C.perfringens agar were also tested. All of these strains were isolated on TSC agar and on CHROMagar™ C.perfringens agar.

The appearance of the characteristic colonies on these 2 media is as follows:

Figure 1:

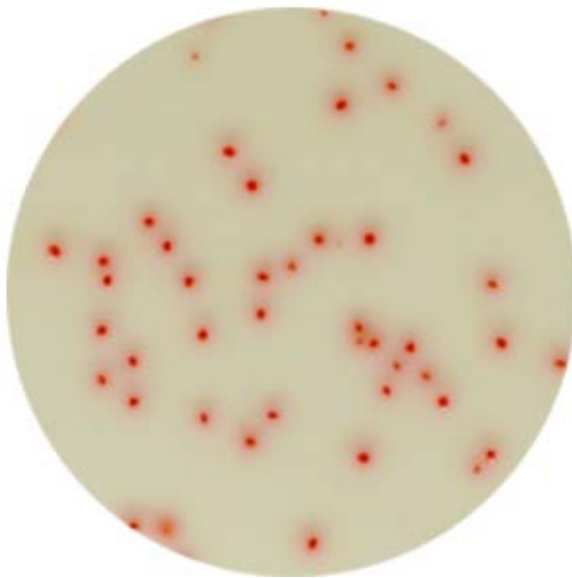
appearance of the characteristic colonies on TSC agar media and CHROMagar™ C.perfringens agar.

TSC agar media



Clostridium perfringens : black colonies

CHROMagar™ C.perfringens agar



Clostridium perfringens : red colonies

Table 1 shows all the strains used for the tests and their aspect on the selective agar media.

Table 1: list of the different bacterial strains isolated on TSC agar media and on CHROMagar™ C. perfringens agar.

| Number | Strain | Origin | TSC | CHROMAGAR |
|--------|---|-----------------------------------|----------------|---------------|
| 1 | <i>Clostridium perfringens</i> | Tap water | Black colonies | Red colonies |
| 3 | <i>Clostridium perfringens</i> | WDCM00007 | Black colonies | Red colonies |
| 6 | <i>Clostridium perfringens</i> | Composite food product | Black colonies | Red colonies |
| 7 | <i>Clostridium perfringens</i> | Spirulina | Black colonies | Red colonies |
| 8 | <i>Clostridium perfringens</i> | Duck meat | Black colonies | Red colonies |
| 9 | <i>Clostridium perfringens</i> | Poultry meat | Black colonies | Red colonies |
| 10 | <i>Clostridium perfringens</i> | Seaweed Dulse | Black colonies | Red colonies |
| 13 | <i>Clostridium perfringens</i> | Sea bean seaweed | Black colonies | Red colonies |
| 14 | <i>Clostridium perfringens</i> | Seaweed Dulse | Black colonies | Red colonies |
| 15 | <i>Clostridium perfringens</i> | Spirulina powder | Black colonies | Red colonies |
| 16 | <i>Clostridium perfringens</i> | Water sewage treatment plant | Black colonies | Red colonies |
| 17 | <i>Clostridium perfringens</i> | Seaweed Laminaria digitata powder | Black colonies | Red colonies |
| 18 | <i>Clostridium perfringens</i> | Water sewage treatment plant | Black colonies | Red colonies |
| 19 | <i>Clostridium perfringens</i> | Pond water | Black colonies | Red colonies |
| 20 | <i>Clostridium perfringens</i> | Seaweed Asco powder | Black colonies | Red colonies |
| 2 | <i>Clostridium butyricum</i> | River water | Black colonies | Red colonies |
| 4 | <i>Clostridium sp.</i> (<i>butyricum</i> or <i>glycolicum</i>) | Outlet water by lagoon treatment | Black colonies | Red colonies |
| 5 | <i>Clostridium glycolicum</i> | Pond water | Black colonies | Red colonies |
| 11 | <i>Clostridium sordelii</i> | Thyme | Black colonies | Red colonies |
| 12 | <i>Clostridium bifermentans</i> | Composite food product | Black colonies | Red colonies |
| N | <i>Paenibacillus sordelii</i> | DSMZ collection | Black colonies | Red colonies |
| P | <i>Clostridium septicum</i> | DSMZ collection | White colonies | No growth |
| A | <i>Leuconostoc mesenteroides</i> | Zucchini flan | No growth | Blue colonies |
| B | <i>Lactobacillus sakei</i> | Smoked salmon | White colonies | Red colonies |
| C | <i>Lactobacillus gasseri</i> | Probiotic | White colonies | Blue colonies |
| D | <i>Lactobacillus reuteri</i> | Probiotic | No growth | Red colonies |
| E | <i>Bifidobacterium lactis</i> | Probiotic | White colonies | Blue colonies |
| F | <i>Lactobacillus paracasei</i> | Dairy product | No growth | No growth |
| G | <i>Leuconostoc mesenteroides</i> | Meat product | No growth | Red colonies |
| H | <i>Streptococcus thermophilus</i> | Probiotic | No growth | Red colonies |
| I | <i>Lactobacillus sakei</i> | Dairy product | White colonies | Red colonies |
| J | <i>Enterococcus faecalis</i> | Bathing water | White colonies | Red colonies |
| K | <i>Enterococcus faecalis</i> | Water sewage treatment plant | White colonies | Red colonies |
| L | <i>Enterococcus faecium</i> | Outlet water by lagoon treatment | White colonies | Red colonies |
| M | <i>Weissella viridescens</i> | Composite food product | No growth | No growth |
| O | <i>Pediococcus pentosaceus</i> | DSMZ collection | No growth | No growth |
| Q | <i>Lactobacillus plantarum</i> | DSMZ collection | No growth | Blue colonies |
| R | <i>Enterobacter cloacae</i> | Outlet water by lagoon treatment | No growth | No growth |
| S | <i>Serratia marcescens</i> | Pond water | No growth | No growth |
| T | <i>Citrobacter freundii</i> | Outlet water by lagoon treatment | No growth | No growth |

2.2.2. Confirmation of *Clostridium perfringens*

2.2.2.1. General

It is recommended, for counts of 1 to 10 colonies, to submit for confirmation all colonies and, for counts greater than 10 colonies, at least 10 colonies randomly chosen.

The ISO 14189:2017 standard indicates that the confirmation of the characteristic colonies on TSC agar must be carried out after subculturing on a blood agar, a Columbia agar or a nutrient-rich agar (for example: Tryptone Soy Agar). These subcultures are then subjected to the acid phosphatase test.

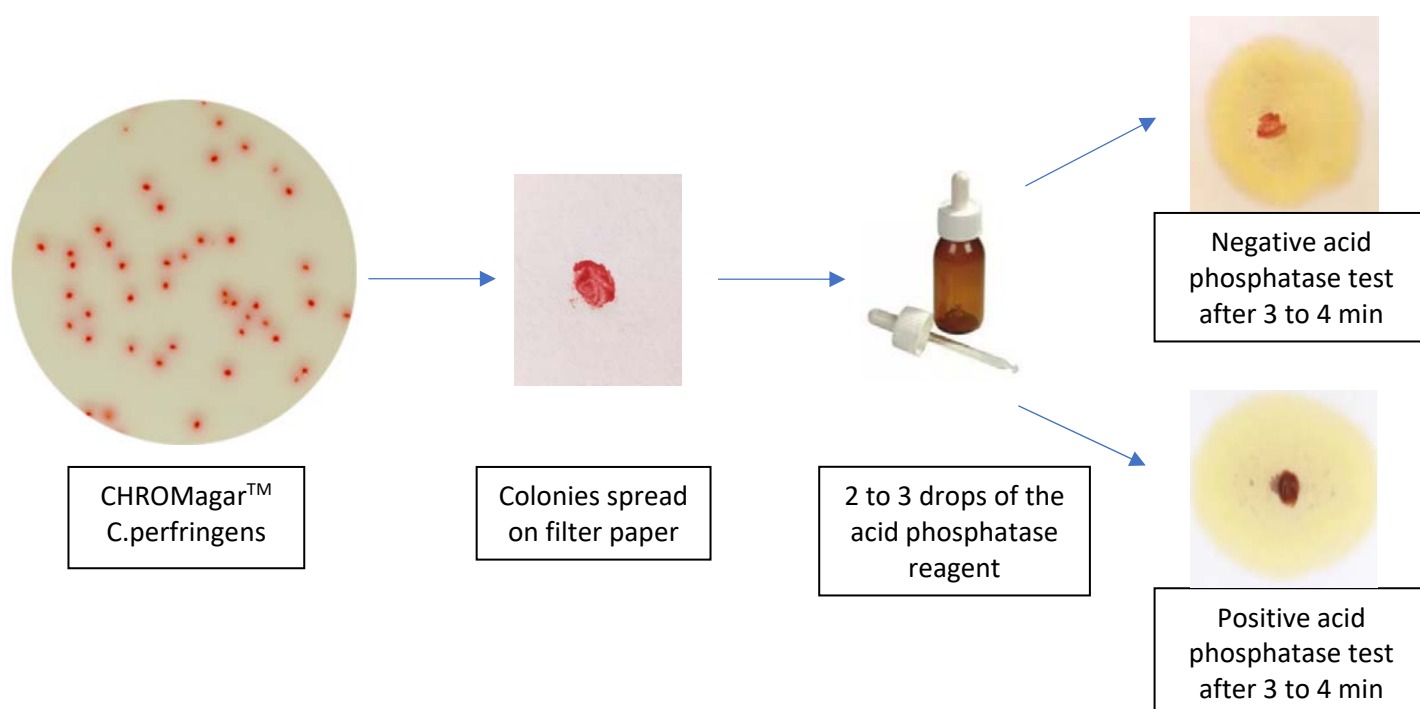
The objective of this study is to determine the effectiveness of the new acid phosphatase reagent directly on the red colonies present on the CHROMagar™ *C.perfringens* agar.

2.2.2.2. Acid phosphatase test

Colonies grown anaerobically are spread on filter paper and 2 to 3 drops of the acid phosphatase reagent are placed onto the colonies. A purplish colour developed within 3 minutes to 4 minutes is considered as a positive reaction. The replacement of the FAST Blue B salt by the FAST Blue RR salt, gives a yellow color to the reagent and allows an easier reading contrast of the positive colonies.

Clostridium perfringens produces black colonies on TSC agar, red colonies on CHROMagar™ *C.perfringens* agar and has an acid phosphatase activity.

Figure 2: principle of the acid phosphatase test



3. Results

3.1. Raw results

All the strains of *Clostridium* for which growth has been observed on CHROMagar™ C.perfringens agar (red colonies or blue colonies) were tested with the acid phosphatase reagent. In parallel, an isolation on a Columbia agar as provided for in the standard ISO 14189:2017 was carried out. An acid phosphatase test (as in ISO 14189:2017 but with FAST Blue RR salt) was then carried out on the colonies present on the Columbia agar. The different results are presented in table 2.

The strains not belonging to the genus *Clostridium* but which may have similar growth characteristics, were isolated on CHROMagar™ C.perfringens agar and tested by the acid phosphatase reagent. In parallel, an isolation on TSA agar was carried out. The results are presented in table 3 and in appendices A and B.

Table 2: results of the acid phosphatase tests on *Clostridium* strains

| Number | Strain | Origin | TSC | CHROMAGAR | | Final results (CHROMAGAR and phosphatase) | COLUMBIA | | Result |
|--------|--|-----------------------------------|----------------|--------------|-----------------------|--|----------------|-------------|---|
| | | | | Aspect | Phosphatase result | | Aspect | Phosphatase | |
| 1 | <i>Clostridium perfringens</i> | Tap water | Black colonies | Red colonies | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 3 | <i>Clostridium perfringens</i> | WDCM00007 | Black colonies | Red colonies | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 6 | <i>Clostridium perfringens</i> | Composite food product | Black colonies | Red colonies | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 7 | <i>Clostridium perfringens</i> | Spirulina | Black colonies | Red colonies | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 8 | <i>Clostridium perfringens</i> | Duck meat | Black colonies | Red colonies | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 9 | <i>Clostridium perfringens</i> | Poultry meat | Black colonies | Red colonies | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 10 | <i>Clostridium perfringens</i> | Seaweed Dulse | Black colonies | Red colonies | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 13 | <i>Clostridium perfringens</i> | Sea bean seaweed | Black colonies | Red colonies | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 14 | <i>Clostridium perfringens</i> | Seaweed Dulse | Black colonies | Red colonies | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 15 | <i>Clostridium perfringens</i> | Spirulina powder | Black colonies | Red colonies | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 16 | <i>Clostridium perfringens</i> | Water sewage treatment plant | Black colonies | Red colonies | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 17 | <i>Clostridium perfringens</i> | Seaweed Laminaria digitata powder | Black colonies | Red colonies | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 18 | <i>Clostridium perfringens</i> | Water sewage treatment plant | Black colonies | Red colonies | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 19 | <i>Clostridium perfringens</i> | Pond water | Black colonies | Red colonies | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 20 | <i>Clostridium perfringens</i> | Seaweed Asco powder | Black colonies | Red colonies | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 2 | <i>Clostridium butyricum</i> | River water | Black colonies | Red colonies | Negative | No <i>Clostridium perfringens</i> | White colonies | Negative | Not <i>Clostridium perfringens</i> |
| 4 | <i>Clostridium</i> sp. (<i>butyricum</i> ou <i>glycolicum</i>) | Outlet water by lagoon treatment | Black colonies | Red colonies | Negative | No <i>Clostridium perfringens</i> | White colonies | Negative | Not <i>Clostridium perfringens</i> |
| 5 | <i>Clostridium glycolicum</i> | Pond water | Black colonies | Red colonies | Negative | No <i>Clostridium perfringens</i> | White colonies | Negative | Not <i>Clostridium perfringens</i> |
| 11 | <i>Clostridium sordelii</i> | Thyme | Black colonies | Red colonies | Negative | No <i>Clostridium perfringens</i> | White colonies | Negative | Not <i>Clostridium perfringens</i> |
| 12 | <i>Clostridium bifermentans</i> | Composite food product | Black colonies | Red colonies | Negative | No <i>Clostridium perfringens</i> | White colonies | Negative | Not <i>Clostridium perfringens</i> |
| N | <i>Paeniclostridium sordelii</i> | DSMZ collection | Black colonies | Red colonies | Negative | No <i>Clostridium perfringens</i> | White colonies | Negative | Not <i>Clostridium perfringens</i> |
| P | <i>Clostridium septicum</i> | DSMZ collection | White colonies | No growth | Ø | No <i>Clostridium perfringens</i> | White colonies | Positive | Possible <i>Clostridium perfringens</i> |

Table 3: results of the acid phosphatase tests on strains not belonging to the genus *Clostridium*.

| Number | Strain | Origin | TSC | CHROMAGAR | | Final result (CHROMAGAR and phosphatase) | TSA | |
|----------|-----------------------------------|----------------------------------|----------------|---------------|-------------|--|----------------|----------------------|
| | | | | Aspect | Phosphatase | | Aspect | Phosphatase |
| A | <i>Leuconostoc mesenteroides</i> | Zucchini flan | No growth | Blue colonies | Negative | No <i>Clostridium perfringens</i> | White colonies | Negative |
| B | <i>Lactobacillus sakei</i> | Smoked salmon | No growth | Red colonies | Positive | Possible <i>Clostridium perfringens</i> | White colonies | Positive |
| C | <i>Lactobacillus gasseri</i> | Probiotic | No growth | Blue colonies | Negative | No <i>Clostridium perfringens</i> | White colonies | Negative |
| D | <i>Lactobacillus reuteri</i> | Probiotic | No growth | Red colonies | Negative | No <i>Clostridium perfringens</i> | White colonies | Negative |
| E | <i>Bifidobacterium lactis</i> | Probiotic | No growth | Blue colonies | Negative | No <i>Clostridium perfringens</i> | White colonies | Negative |
| F | <i>Lactobacillus paracasei</i> | Dairy product | No growth | No growth | Ø | No <i>Clostridium perfringens</i> | White colonies | Positive (very weak) |
| G | <i>Leuconostoc mesenteroides</i> | Meat product | No growth | Red colonies | Negative | No <i>Clostridium perfringens</i> | White colonies | Negative |
| H | <i>Streptococcus thermophilus</i> | Probiotic | No growth | Red colonies | Negative | No <i>Clostridium perfringens</i> | White colonies | Negative |
| I | <i>Lactobacillus sakei</i> | Dairy product | No growth | Red colonies | Negative | No <i>Clostridium perfringens</i> | White colonies | Negative |
| J | <i>Enterococcus faecalis</i> | Bathing water | White colonies | Red colonies | Negative | No <i>Clostridium perfringens</i> | White colonies | Negative |
| K | <i>Enterococcus faecalis</i> | Water sewage treatment plant | White colonies | Red colonies | Negative | No <i>Clostridium perfringens</i> | White colonies | Negative |
| L | <i>Enterococcus faecium</i> | Outlet water by lagoon treatment | White colonies | Red colonies | Negative | No <i>Clostridium perfringens</i> | White colonies | Negative |
| M | <i>Weissella viridescens</i> | Composite food product | No growth | No growth | Ø | No <i>Clostridium perfringens</i> | White colonies | Negative |
| O | <i>Pediococcus pentosaceus</i> | DSMZ collection | No growth | No growth | Ø | No <i>Clostridium perfringens</i> | White colonies | Negative |
| Q | <i>Lactobacillus plantarum</i> | DSMZ collection | No growth | Blue colonies | Negative | No <i>Clostridium perfringens</i> | White colonies | Negative |
| R | <i>Enterobacter cloacae</i> | Outlet water by lagoon treatment | White colonies | No growth | Ø | No <i>Clostridium perfringens</i> | White colonies | Negative |
| S | <i>Serratia marescens</i> | Pond water | No growth | No growth | Ø | No <i>Clostridium perfringens</i> | White colonies | Positive |
| T | <i>Citrobacter freundii</i> | Outlet water by lagoon treatment | No growth | No growth | Ø | No <i>Clostridium perfringens</i> | White colonies | Positive |

3.2. Comments

22 strains belonging to the genus *Clostridium* and 18 other strains of different genera were tested during this study. 21 strains out of the 22 *Clostridium* strains developed characteristic red colonies on CHROMagar™ C.perfringens agar medium and 15 of these strains gave a positive reaction to the modified acid phosphatase test, both from CHROMagar™ C.perfringens agar and COLUMBIA agar. All the strains of *Clostridium* spp. exclusive of *Clostridium perfringens*, gave a negative result with the acid phosphatase test.

Among the 18 strains of bacteria not belonging to the genus *Clostridium*, 8 of them gave characteristic colonies on the CHROMagar™ C.perfringens agar medium:

- Strain B: *Lactobacillus sakei*
- Strain D: *Lactobacillus reuteri*
- Strain G: *Leuconostoc mesenteroides*
- Strain H: *Streptococcus thermophilus*
- Strain I: *Lactobacillus sakei*
- Strain J: *Enterococcus faecalis*
- Strain K: *Enterococcus faecalis*
- Strain L: *Enterococcus faecium*

For all these strains except for *Lactobacillus sakei* (B), the acid phosphatase test was negative. A second strain of *Lactobacillus sakei* (I) from a different origin has been tested and gave a negative result.

4. Interpretation and conclusion

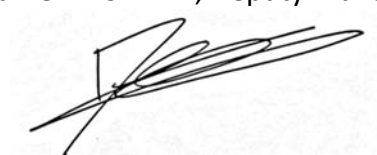
The present study involved 40 strains analyzed with the CHROMagar™ C.perfringens agar media and confirmed with the modified acid phosphatase test. No difference was observed between the confirmation results by the modified acid phosphatase test carried out directly from the CHROMagar™ C.perfringens agar or after purification on Columbia or TSA. A single strain of *Lactobacillus sakei* gave a false positive result, another strain of *Lactobacillus sakei* having given a negative result on both CHROMagar™ C.perfringens and Columbia agar media.

Particular attention must be paid when reading the acid phosphatase test when it is carried out directly from the red colonies on CHROMagar™ C.perfringens agar. Indeed, the appearance of a purple coloration for a positive sample is obtained after 3 to 4 minutes and a training or a warning specific to the operator must be specified to avoid the false positive results of a red colony.

In conclusion, it appears that the replacement of the FAST Blue B salt by the FAST Blue RR salt has no impact on the results obtained with the CHROMagar™ C.perfringens agar media and that the reading contrast is satisfactory for the detection of *Clostridium perfringens*.

This conclusion is only valid in the operating conditions of the study and with the bacterial strains tested.








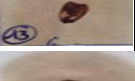





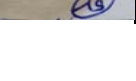
Le Lion d'Angers, May 25, 2020
Guillaume MESNARD, Deputy Manager










APPENDICES

Raw results of study





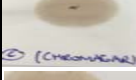





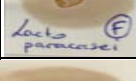




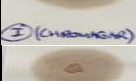

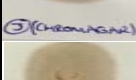





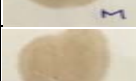

APPENDIX A: Raw results for strains of *Clostridium*

| Number | Strain (identified by MALDI-TOF) | Origin | TSC | CHROMAGAR | | | Final result (CHROMAGAR and phosphatase) | COLUMBIA | | Result |
|--------|-------------------------------------|-----------------------------------|----------------|--------------|---|--------------------|--|----------------|-------------|--------------------------------|
| | | | | Aspect | Phosphatase | Phosphatase result | | Aspect | Phosphatase | |
| 1 | <i>Clostridium perfringens</i> | Tap water | Black colonies | Red colonies |  | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 3 | <i>Clostridium perfringens</i> | WDCM00007 | Black colonies | Red colonies |  | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 6 | <i>Clostridium perfringens</i> | Composite food product | Black colonies | Red colonies |  | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 7 | <i>Clostridium perfringens</i> | Spirulina | Black colonies | Red colonies |  | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 8 | <i>Clostridium perfringens</i> | Duck meat | Black colonies | Red colonies |  | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 9 | <i>Clostridium perfringens</i> | Poultry meat | Black colonies | Red colonies |  | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 10 | <i>Clostridium perfringens</i> | Seaweed Dulse | Black colonies | Red colonies |  | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 13 | <i>Clostridium perfringens</i> | Sea bean seaweed | Black colonies | Red colonies |  | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 14 | <i>Clostridium perfringens</i> | Seaweed Dulse | Black colonies | Red colonies |  | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 15 | <i>Clostridium perfringens</i> | Spirulina powder | Black colonies | Red colonies |  | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 16 | <i>Clostridium perfringens</i> | Water sewage treatment plant | Black colonies | Red colonies |  | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 17 | <i>Clostridium perfringens</i> | Seaweed Laminaria digitata powder | Black colonies | Red colonies |  | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 18 | <i>Clostridium perfringens</i> | Water sewage treatment plant | Black colonies | Red colonies |  | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 19 | <i>Clostridium perfringens</i> | Pond water | Black colonies | Red colonies |  | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |




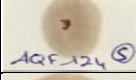

APPENDIX A: Raw results for strains of *Clostridium*

| Number | Strain (identified by MALDI-TOF) | Origin | TSC | CHROMAGAR | | | Final result (CHROMAGAR and phosphatase) | COLUMBIA | | Result |
|--------|--|----------------------------------|----------------|--------------|---|-----------------------|---|----------------|-------------|---|
| | | | | Aspect | Phosphatase | Phosphatase result | | Aspect | Phosphatase | |
| 20 | <i>Clostridium perfringens</i> | Seaweed Asco powder | Black colonies | Red colonies |  | Positive | <i>Clostridium perfringens</i> | White colonies | Positive | <i>Clostridium perfringens</i> |
| 2 | <i>Clostridium butyricum</i> | River water | Black colonies | Red colonies |  | Negative | Not <i>Clostridium perfringens</i> | White colonies | Negative | Not <i>Clostridium perfringens</i> |
| 4 | <i>Clostridium</i> sp. (<i>butyricum</i> or <i>glycolicum</i>) | Outlet water by lagoon treatment | Black colonies | Red colonies |  | Negative | Not <i>Clostridium perfringens</i> | White colonies | Negative | Not <i>Clostridium perfringens</i> |
| 5 | <i>Clostridium glycolicum</i> | Pond water | Black colonies | Red colonies |  | Negative | Not <i>Clostridium perfringens</i> | White colonies | Negative | Not <i>Clostridium perfringens</i> |
| 11 | <i>Clostridium sordelii</i> | Thyme | Black colonies | Red colonies |  | Negative | Not <i>Clostridium perfringens</i> | White colonies | Negative | Not <i>Clostridium perfringens</i> |
| 12 | <i>Clostridium bifermentans</i> | Composite food product | Black colonies | Red colonies |  | Negative | Not <i>Clostridium perfringens</i> | White colonies | Negative | Not <i>Clostridium perfringens</i> |
| N | <i>Paenibacillus sordelii</i> | DSMZ collection | Black colonies | Red colonies |  | Negative | Not <i>Clostridium perfringens</i> | White colonies | Negative | Not <i>Clostridium perfringens</i> |
| P | <i>Clostridium septicum</i> | DSMZ collection | White colonies | No growth | Ø | Ø | Not <i>Clostridium perfringens</i> | White colonies | Positive | Possible <i>Clostridium perfringens</i> |

APPENDIX B: Raw results for strains other than *Clostridium*

| Number | Strain (identified by MALDI-TOF) | Origin | TSC | CHROMAGAR | | | Final result (CHROMAGAR and phosphatase) | TSA | | |
|--------|-------------------------------------|----------------------------------|----------------|---------------|---|--------------------|--|----------------|---|--|
| | | | | Aspect | Phosphatase | Phosphatase result | | Aspect | Phosphatase | Phosphatase result |
| A | <i>Leuconostoc mesenteroides</i> | Zucchini flan | No growth | Blue colonies |  | Negative | Not <i>Clostridium perfringens</i> | White colonies |  | Negative |
| B | <i>Lactobacillus sakei</i> | Smoked salmon | No growth | Red colonies |  | Positive | Possible <i>Clostridium perfringens</i> | White colonies |  | Positive |
| C | <i>Lactobacillus gasseri</i> | Probiotic | No growth | Blue colonies |  | Negative | Not <i>Clostridium perfringens</i> | White colonies |  | Negative |
| D | <i>Lactobacillus reuteri</i> | Probiotic | No growth | Red colonies |  | Negative | Not <i>Clostridium perfringens</i> | White colonies |  | Negative |
| E | <i>Bifidobacterium lactis</i> | Probiotic | No growth | Blue colonies |  | Negative | Not <i>Clostridium perfringens</i> | White colonies |  | Negative |
| F | <i>Lactobacillus paracasei</i> | Dairy product | No growth | No growth | Ø | Ø | Not <i>Clostridium perfringens</i> | White colonies |  | Positive (very weak) |
| G | <i>Leuconostoc mesenteroides</i> | Meat product | No growth | Red colonies |  | Negative | Not <i>Clostridium perfringens</i> | White colonies |  | Negative |
| H | <i>Streptococcus thermophilus</i> | Probiotic | No growth | Red colonies |  | Negative | Not <i>Clostridium perfringens</i> | White colonies |  | Negative |
| I | <i>Lactobacillus sakei</i> | Dairy product | No growth | Red colonies |  | Negative | Not <i>Clostridium perfringens</i> | White colonies |  | Negative (weak coloration after 4 min) |
| J | <i>Enterococcus faecalis</i> | Bathing water | White colonies | Red colonies |  | Negative | Not <i>Clostridium perfringens</i> | White colonies |  | Negative (weak coloration after 4 min) |
| K | <i>Enterococcus faecalis</i> | Water sewage treatment plant | White colonies | Red colonies |  | Negative | Not <i>Clostridium perfringens</i> | White colonies |  | Negative |
| L | <i>Enterococcus faecium</i> | Outlet water by lagoon treatment | White colonies | Red colonies |  | Negative | Not <i>Clostridium perfringens</i> | White colonies |  | Negative |
| M | <i>Weissella viridescens</i> | Composite food product | No growth | No growth | Ø | Ø | Not <i>Clostridium perfringens</i> | White colonies |  | Negative |
| O | <i>Pediococcus pentosaceus</i> | DSMZ collection | No growth | No growth | Ø | Ø | Not <i>Clostridium perfringens</i> | White colonies |  | Negative |

APPENDIX B: Raw results for strains other than *Clostridium*

| Number | Strain (identified by MALDI-TOF) | Origin | TSC | CHROMAGAR | | | Final result (CHROMAGAR and phosphatase) | TSA | | |
|--------|-------------------------------------|----------------------------------|----------------|---------------|---|-----------------------|---|----------------|---|-----------------------|
| | | | | Aspect | Phosphatase | Phosphatase result | | Aspect | Phosphatase | Phosphatase result |
| Q | <i>Lactobacillus plantarum</i> | DSMZ collection | No growth | Blue colonies |  | Negative | Not <i>Clostridium perfringens</i> | White colonies |  | Negative |
| R | <i>Enterobacter cloacae</i> | Outlet water by lagoon treatment | White colonies | No growth | Ø | Ø | Not <i>Clostridium perfringens</i> | White colonies |  | Negative |
| S | <i>Serratia marcescens</i> | Pond water | No growth | No growth | Ø | Ø | Not <i>Clostridium perfringens</i> | White colonies |  | Positive |
| T | <i>Citrobacter freundii</i> | Outlet water by lagoon treatment | No growth | No growth | Ø | Ø | Not <i>Clostridium perfringens</i> | White colonies |  | Positive |