

Preliminary study on enumeration medium of presumptive Bacillus cereus

Preliminary Report

Réf. Proposition : SBM n° 10.845.3

I. INTRODUCTION

The company CHROMagar requests ADRIA NORMANDIE to study an enumeration medium of presumptive Bacillus cereus.

II. TECHNICAL APPROACH

The proposed technical approach is a study of the selectivity of the enumeration medium of presumptive CHROMagar B.cereus as well as an application on said medium on potentially naturally or artificially contaminated samples.

II. 1 Study of the selectivity

The purpose of this step is to study the selectivity of CHROMagar B. cereus. To do this, inclusivity and exclusivity of the medium were studied. For inclusivity, 30 pure strains of B. cereus were isolated on the medium to ensure that they have distinctive morphological characteristics. These strains were selected from the collection of ADRIA Normandie to present the most biodiversity in terms of genetic (presence of 7 groups) and origin (products). For exclusivity, 20 pure strains of genus or species close to to *B. cereus* were isolated on the medium to ensure that they do not have the distinctive morphological characteristics of *B. cereus*. These strains correspond to gender and species commonly found in food products (for example: *B. subtilis, B. licheniformis, B. megaterium, B. amyloliquefaciens, B. simplex, Paenibacillus spp. Sporosarcina spp. Viridibacillus spp.*, ...) but also to the B. mycoides species in order to establish whether the medium is specific for the species *B. cereus* group or to cereus group.

II. 2 Analysis of potentially naturally contaminated samples

The technical approach proposed is a comparison of enumerations obtained on CHROMagar medium for presumptive B. cereus with those obtained on the Mossel medium (AES) on 20 samples likely to have a natural contamination of presumptive *B. cereus* because of the nature of the selected food matrices. Counts were performed according to standard NF EN ISO 7932-July 2005 with reading at 24 and 48 hours on both media. Confirmations were made by hemolysis test with sheep blood agar on one single typical colony (note: the standard 7932 application ask for 5 colonies for confirmation) after overnight incubation at 30°C.

II. 3 Analysis of artificially contaminated samples

The technical approach proposed is a comparison of enumerations obtained on CHROMagar medium for presumptive *B. cereus* with those obtained on the Mossel medium (AES) on 3 food matrices (dairy products, surimi and mashed vegetables) artificially contaminated with one mesophilic strain of *B. cereus* (vegetative form) at different rates (10, 100, 1000 CFU/g) with 3 replications. Analyses were performed after 1h at 4°C (time to adapt to the matrix). The analyses were performed according to standard NF EN ISO 7932-July 2005 with reading at 24 and 48 h on both media.

Note: In all cases, products from CHROMagar method tested were given free of charge by CHROMagar Company.

III. RESULTS

III. 1 Study of the selectivity

The detailed results are provided in Appendix 1

III.1.1 Inclusivity study

Thirty pure strains of presumptive *Bacillus cereus* were selected with respect to their origin or genetic groups: 15 strains from dairy products, surimi, 4, 5 egg products, 2 and 4 surface samples are strains of collection. These strains are divided into several genetic groups (group II to group VII). 30 isolates (except one confirmed non *B. cereus*) have the expected characteristics of the medium tested namely a turquoise color more or less intense, with a presence of a halo around the colonies. However, we can distinguish three groups of colonies relative to the observed macroscopic appearance:

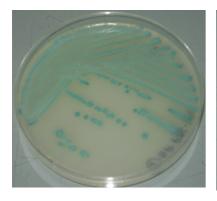
- Group 1: represents the largest group with presence after 24 h of incubation at 30 ° C in small colonies (1-2 mm diameter), regular and intense turquoise color



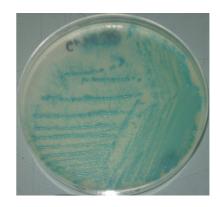
- Group 2: colonies of 3 to 5 mm in diameter after 24 h of incubation at 30 ° C



- Group 3: different types of colonies are represented in this group: less intense color (blue-green) with the presence of a halo less marked, irregular colonies and colonies characteristic of Bacillus mycoides







Very intense color, reduced halo

regular colonies

Bacillus mycoides

After 48 h of incubation, the same features are found with some drying visible colonies or a slight increase in the diameter of others.

In all cases, the CHROMagar B. cereus has allowed the identification of typical colonies of B. cereus and B. mycoides. From macroscopic observations we can conclude that the medium is discriminating after 24 hours of incubation at 30 ° C.

III.1.2 Exclusivity Study

Twenty pure strains of genus and species close to *B. cereus* were isolated on CHROMagar B. cereus medium. Observation of the potential growth and the macroscopic appearance was performed for all selected strains. The results of macroscopic observations showed that growth was inhibited for most strains tested. When growth was observed (after 24h or 48h), it was not characteristic of *B. cereus* (turquoise blue colonies without halo or white or pinkish).

Conclusions about the selectivity of the medium

The study of the selectivity of the medium CHROMagar B. cereus showed the usefulness of the medium to identify strains belonging to B. cereus group (at least *B. cereus* and *B. mycoides*). Of the 50 pure strains tested, no false-positive or false-negative was observed under the conditions used. In addition, results were obtained after 24 h of incubation. Finally, no concern for practicability of use of the medium was observed.

Appendix 1

Step 1 Bacillus cereus studied strains

	CODE	GROUP	ORIGIN	Appearance at 24H		Appearance at 48H		
				color	growth	color	growth	
1	8P P26-44	II	milk powder	pale	+++, localised halo	same	same	
2	A7 ovo10	II	Egg product	Nothing to say	+++, 2mm	same	dry colonies with white outline	
3	AS 11 ovo14	II	Egg product	Nothing to say	+++, 2mm	same	dry colonies with white outline	
4	44009	II	Plumber	Nothing to say	+++, 2mm	same	dry colonies with white outline	
5	29014	П	floating islands	Nothing to say	+++, 2mm	same	dry colonies with white outline	
6	28056	II	Rum/grapes	Nothing to say	+++, 2mm	same	dry colonies with white outline	
7	27009Vg	II	Egg cream	Nothing to say	+++, 2mm	same	dry colonies with white outline	
8	12P P26-52	III	milk powder	Nothing to say	+++, 2mm	same	same	
9	15001 22	III	surimi	Nothing to say	+++, 2mm	same	same	
10	33007	III	Milk product	Nothing to say	+++, 2mm	same	same	
11	28040	III	coconut	Less intense Nothing to	+++, 4mm irregular	same	same	
12	INRA 91	III	collection	say	+++, 4 mm	same	same	
13	13049-22	Ш	surimi	No growth : confirmed strain non <i>B.cereus</i>				
14	B1 ovo 20	IV	Egg product	Nothing to say	+++, 2mm	same	same	
15	07015-2	IV	surimi	Nothing to say	+++, 2mm	same	same	

Appendix 1 (following table, step 1)

	CODE	GROUP	ORIGIN	Арре	earance at 24H	Appearance at 48H		
	CODE			color	growth	color	growth	
16	49006 RVg	IV	Panacotta	Nothing to say	+++, 2mm	same	Dry colonies with white outline	
17	46001 Msp	IV	Egg waste before heating	Nothing to say	+++, 2mm	same	4mm colonies	
18	51007 Rsp	IV	Egg waste after heating	Nothing to say	+++, 2mm	same	same	
19	5012	V	Floating islands	Nothing to say	+++, 2mm	same	same	
20	AS 44-1	V	product testing witness	Nothing to say	+++, 2mm	same	same	
21	AS 27-1	V	Milk product	Nothing to say	+++, 2mm	same	same	
22	AS 36	V	Milk product	Nothing to say	+++, 2mm	same	same	
23	42007 RVg	V	Panacotta	Nothing to say	+++, 2mm	same	same	
24	KBAB4	VI	collection	Nothing to say	+++, 2mm	same	same	
25	13P' P26- 54''	VI	Milk powder	Nothing to say	+++, 2mm	same	4mm colonies	
26	A11 I Mossel	VI	Surface sample	Nothing to say	+++, 2mm	same	same	
27	D2PPC	VI	Milk product CAR coffee	Nothing to say	+++, 2mm	same	4mm colonies	
28	51,19	VI	collection	Nathinata	B.mycoides			
29	D19	VII	collection	Nothing to say	+++,4mm	same	same	
30	13051-1	?	surimi	Nothing to say	+++,4mm	same	same	

non-Bacillus cereus studied strains

	CODE	GROUP	ORIGIN	Appearance a	at 24H	Appearance at 48H		
	CODE	GROUP		color	growth	color	growth	
Α	LMG 19409	B. licheniformis	?	/	No growth	/	same	
В	Ad 978	B. licheniformis	collection	Blue without halo uncharacteristic	+	same	same	
С	1003006 2 FAM	B. licheniformis	rice flour	Blue without halo uncharacteristic	+	same	same	
D	16001-22	B. subtilis	surimi	,	No growth	+/- blue uncharacteristic	+	
				Blue without halo				
E	932001	B. subtilis	Milk dessert	uncharacteristic	+	same	same	
F	1040008	B. circulans	Chocolate- orange dose	Blue without halo uncharacteristic	+	same	same	
G	1001003 aé spo	B. subtilis	rice flour	Blue without halo uncharacteristic	+	same	same	
Н	1042001	B. subtilis	dessert caramel cream	Blue without halo uncharacteristic	+	same	same	
ı	1033012	B. megaterium	wipe VP	Blue without halo uncharacteristic	+	same	same	
j	114 10	B. amyloliquefaciens	rice pudding	Blue without halo uncharacteristic	+	same	same	
			nce padding	Blue without halo				
K	B Nov 42037	B. amyloliquefaciens	shredded	uncharacteristic	+	same	same	
L	18022-3	B. simplex	surimi	/	No growth	white	+	
М	1010003	P. lautus	Egg product	/	No growth	/	same	
N	1009001 majo	P. spp	chocolate	Blue without halo uncharacteristic	+	same	same	
0	1023015 FAM	P. spp	Mashed potatoes	Blue without halo uncharacteristic	+	same	same	
Р	1007001 PC	Sp. aquimarina	Chocolat cream	White uncharacteristic	+++	Pinkish uncharacteristic	same	
Q	17033-4	Sp. aquimarina	shredded surimi	White uncharacteristic	+++	pinkish	same	
R	0915004 PCA	B. pumilus	shredded 3 chocolates	/	No growth	White uncharacteristic	+	
11	1001002 aé	D. parimus	chocolates	<i>1</i>	140 810 4/(11	White	•	
S	spo	B. pumilus	rice	/	No growth	uncharacteristic	+	
Т	3040053	B. circulans	Milky pudding	Blue without halo uncharacteristic	+	same	same	